## **ORIGINAL RESEARCH PAPER**

# EFFECT OF AMINO ACIDS ADDED TO CULTURE MEDIUM ON THE GROWTH AND SURVIVAL OF *LACTOBACILLUS BULGARICUS LB6* DURING FREEZE-DRYING

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Plackett–Burman design was applied to evaluate the effects of amino acids on the growth in medium and survival during freeze-drying of *Lactobacillus bulgaricus* LB6. Moreover, in consideration of the optimal amino acids for the growth and survival of *Lactobacillus bulgaricus* LB6 before and after freeze-drying, viable counts and survival rate were monitored in the medium containing selected amino acids (Glutamate, Alanine, Glycine, Leucine, Serine, Arginine, Lysine and Hydroxyproline). The results indicated that Leucine (p=0.0117 for viable counts, p=0.0121 for survival rate) and Arginine (p=0.0120 for viable counts, p=0.0043 for survival rate) out of the investigated amino acids can significantly affect both the growth and survival rate of *Lactobacillus bulgaricus LB*6, and have a positive effect.

Keywords: Lactobacillus bulgaricus, promoting-growth, freeze-drying, amino acid

## Introduction

Dairy starter cultures are of industrial importance and commercial significance for fermented foods, and have been well recognized worldwide (Carvalho *et al.*, 2004b; Santivarangkna *et al.*, 2008). The *Lactobacillus bulgaricus*, a lactic acid bacterium,

is able to produce lactic acid in the production of yogurt, cheese and other fermented products (Guchte *et al.*, 2006), and is of vital importance to the fermented food in combination with *Streptococcus thermophilus*.

The efficacy of L. bulgaricus as starter cultures for the dairy industry depends strongly on the number of viable and active cells (Chen et al., 2014). Thus, during the preparation of the starter cultures, the production and maintenance techniques that maximize the viability and activity of the bacterial cells must be established. The lactic acid bacterium had been preserved and distributed in several forms, such as liquid, spray-dried and lyophilised. The lyophilized or freeze-drying is the most convenient and successful method of preserving bacteria (Berny and Hennebert, 1991), and has been widely used in microbiology for many decades to stabilize and store cultures (Morgan and Vesey, 2009). However, not all cells survived this treatment and a survival rate as low as 0.1% has been reported (Abadias et al., 2001). The major causes of cell viability loss during freeze-drying are related to ice crystal formation, membrane damage from high osmolarity due to high concentrations of internal solutes, maromolecule denaturation, and the removal of water, which affects properties of many hydrophilic macromolecules in cells (Fonseca et al., 2000; Fowler and Toner, 2005; Brennan et al., 1986; Allison et al., 1999). Thus, to protect the viability of probiotics during dehydration, people have added varieties of protective agents to the drying media before freeze-drying (Hubalek, 2003). The carbohydrates that have protective effects for probiotic bacteria during freeze-drying were well documented, for instance, sorbitol (Linders et al., 1997a; Foerst et al., 2012), mannitol (Efiuvwevwere et al., 1999), sucrose (Carvalho et al., 2003a), lactose (Higl et al., 2007), and mannose (Carvalho et al., 2004a), inulin and fructooligosaccharides (Clarissa et al., 2007). Amino acids, including phenylalanine, arginine, glycine (Mattern et al., 1999) and sodium glutamate (Font et al., 1983; Teixeira et al., 1995) were employed to protect the cells. Some salt buffers, such as NaCl or KCl (Carvalho et al., 2003a), sodium citrate (Kets et al., 2004; Lone et al., 2009), phosphate (Ohtake, 2004), calcium carbonate and manganese sulphate can help to protect cells during freeze-drying together with other protectants.

On the other hand, it is well known that the growth of bacterial cultures varies depending on the growth medium, and the composition of the growth media as a contributing factor to the survival rate of probiotic cultures during drying has been demonstrated (Meng *et al.*, 2008). The presence of sugars, such as lactose, sucrose, trehalose, mannose, fructose, glucose, fructose etc. in the growth media has an impact on the survival rate of probiotic cultures during drying (Ferreira *et al.*, 2005;

Carvalho *et al.*, 2003b; Carvalho *et al.*, 2004a). Other additives that can affect the viability or survival rate used in growth media were NaCl (Linders *et al.*, 1997b), manganese sulphate, Tween 80 and ascorbic acid (Carvalho *et al.*, 2003b), carnitine and betaine (Kets and de Bont, 1994; Åsa *et al.*, 2012). There is still lack of studies on the influence of growth media on the subsequent survival of cells during freezedrying, in particularly the insights into the amino acids that can improve the proliferation and survival when added in the growth medium are few. The effects of sugar alcohol and proteins on the survival of *Lactobacillus bulgaricus* LB6 during freezedrying were also studied (Chen *et al.*, 2015).

The aim of the present study was to investigate the potential of different amino acids, added into the culture medium, to act as growth promoting substances for *L*. *bulgaricus LB6*, and protective agents for freeze-drying applications.

### Materials and methods

#### **Microorganisms**

*L. bulgaricus LB6* was obtained from School of Food and Biological Engineering, Shaanxi University of Science & Technology (Xi'an, China), and activated for 24 h at 37 °C with basal LAB growth medium which was repeated three times until the viable counts were stable. The basal LAB growth medium contained 20g of glucose, 4g yeast extract powder, 10g soya peptone, 1000mL water, which was obtained from Beijing Land Bridge Technology Co., Ltd. MRS medium. The amino acids were added to the basal LAB growth media that were autoclaved and cooled to 50 °C, and 3% active culture was inoculated into the medium and incubated at 37 °C, and then viable counts at 18-20h were performed. All the amino acids were sterilized using 0.22  $\mu$ m membrane filtration before added into the autoclaved medium.

## Vacuum freeze-drying

After incubation, *L. bulgaricus LB6* culture was centrifuged at  $10000 \times g$  for 15min and the supernatant was discarded to harvest *L. bulgaricus LB6* cells. The cells were pre-frozen at -80 °C for 6-12h after protective agents (phosphate buffer) were added, and then frozen at -55 °C, 6.93pa for 24h using a vacuum freeze dryer LGJ-22D (Beijing Four-Ring Science Instrument Plant Co., Ltd., Beijing, China).

### Determination of cell counts

After a serial dilution on sterile saline solution (NaCl, 0.9% w/v), the diluted bacterial suspension was aliquoted into 0.1mL doses with Hamilton syringe and dropped into a count plate, then spread uniformly. The count plates were incubated

at 37°C anaerobically for 36-48h and then the viable *L. bulgaricus* cells were counted (Shu *et al.*, 2014). The freeze-dried powders were reconstituted to their original prefreeze dried volumes by adding sterile saline solution and the number of viable cells counted as above.

## Calculation of survival

Survival percentage was calculated as the number of viable cells after drying/number of viable cells before drying×100%.

### Experimental Design of Amino Acid Screening

The goal of applying Plackett–Burman design was to identify which factors of the selected amino acids (Glutamate, Alanine, Glycine, Leucine, Serine, Arginine, Lysine and Hydroxyproline) have significant effect on both viable counts and survival rate before and after freeze-drying. According to Plackett–Burman design, all eight factors were tested at a lower and a higher level coded as (+1) and (-1) (Table 1), respectively. The design matrix is shown in Table 2 where can be seen the effect of the 11 variables (including three error terms: Lactose (X2), Fructooligosaccharides (X10) and Galactooligosaccharides (X11), in order to estimate the standard deviation) as resulted after running 12 independent experimental tests.

**Table 1.** Amino acids tested in a Plackett-Burman survey for their efficacy in increasing the viability and cell survival of *Lactobacillus bulgaricus* during freeze-drying

| Variables | Medium components | Lower level (mg/L) | Higher level (mg/L) |  |
|-----------|-------------------|--------------------|---------------------|--|
| X1        | Glutamate         | 4                  | 6                   |  |
| X3        | Alanine           | 4                  | 6                   |  |
| X4        | Glycine           | 4                  | 6                   |  |
| X5        | Leucine           | 4                  | 6                   |  |
| X6        | Serine            | 4                  | 6                   |  |
| X7        | Arginine          | 4                  | 6                   |  |
| X8        | Lysine            | 4                  | 6                   |  |
| X9        | Hydroxyproline    | 4                  | 6                   |  |

The statistical analysis was performed by the Design-Expert (Version, 8.0.6) to identify the significant variables and their corresponding coefficients, so that the levels of various can be managed to obtain a desired output. Hence, F-value, sum of squares, p-value and confidence interval (CI) were analyzed using the experimental

results of the viable bacteria and survival rate. The experimental results (response function, Y) were fitted to first order multiple regression equations (Eq. (1)) using the coded level (-1 or +1) of the variables (Xi):

$$Y = b_{o} + \sum_{i=1}^{k} b_{i} x_{i}$$
(1)

## Results

#### The experimental design and results

In the present study, the experimental design and results showed in Table 2 are followed by the Plackett–Burman design. The value Y1 stands for viable counts before centrifuged (the unit  $10^9$  CFU/mL) and Y2 (%) for survival rate after freeze-drying.

 Table 2. The Plackett-Burman experimental design matrix and results for the evaluated data

| Ru          | X1 | X2 | X3 | X  | X  | X  | X  | X    | X    | X1 | Х  | Y1/10 <sup>9</sup> CFU/ | Y2    |
|-------------|----|----|----|----|----|----|----|------|------|----|----|-------------------------|-------|
| 1           | 1  | -1 | 1  | -1 | -1 | -1 | 1  | 1    | 1    | -1 | 1  | 1.70                    | 7.35  |
| 2           | 1  | 1  | -1 | 1  | -1 | -1 | -1 | 1    | 1    | 1  | -1 | 1.20                    | 23.00 |
| 3           | -1 | 1  | 1  | -1 | 1  | -1 | -1 | -1   | 1    | 1  | 1  | 1.66                    | 8.07  |
| 4           | 1  | -1 | 1  | 1  | -1 | 1  | -1 | -1   | -1   | 1  | 1  | 1.74                    | 15.11 |
| 5           | 1  | 1  | -1 | 1  | 1  | -1 | 1  | -1   | -1   | -1 | 1  | 9.45                    | 1.66  |
| 6           | 1  | 1  | 1  | -1 | 1  | 1  | -1 | 1    | -1   | -1 | -1 | 1.93                    | 6.94  |
| 7           | -1 | 1  | 1  | 1  | -1 | 1  | 1  | -1   | 1    | -1 | -1 | 1.06                    | 10.85 |
| 8           | -1 | -1 | 1  | 1  | 1  | -1 | 1  | 1    | -1   | 1  | -1 | 1.38                    | 4.96  |
| 9           | -1 | -1 | -1 | 1  | 1  | 1  | -1 | 1    | 1    | -1 | 1  | 1.59                    | 13.08 |
| 10          | 1  | -1 | -1 | -1 | 1  | 1  | 1  | -1   | 1    | 1  | -1 | 13.5                    | 1.17  |
| 11          | -1 | 1  | -1 | -1 | -1 | 1  | 1  | 1    | -1   | 1  | 1  | 2.33                    | 2.68  |
| 12          | -1 | -1 | -1 | -1 | -1 | -1 | -1 | -1   | -1   | -1 | -1 | 1.42                    | 9.51  |
| The control |    |    |    |    |    |    |    | 0.97 | 0.77 |    |    |                         |       |

### Effect of amino acids on the growth of L. bulgaricus

The analysis of variance (ANOVA) was performed to estimate the effect on the growth of each factor (Table 3). The Model F-value of 19.22 implies the model is significant. The p-value of linear model was 0.0168, which demonstrated that the fit of the linear model was satisfied. Values of "Prob > F" less than 0.0500 indicate

model terms are significant; therefore, the sum of squares confirmed the significances of each amino acid. In this case, Glutamate (X1) (p=0.0116), Alanine (X3) (p=0.0117), Leucine (X5) (p=0.0117), Arginine (X7) (p=0.0120) and Lysine (X8) (p=0.0142) are significant model terms and according to this assumption these five amino acids were found to be significant factors for the growth of *L. bulgaricus*. Values greater than 0.1000 indicate the model terms are not significant. Furthermore, the positive or negative of coefficients in Final Equation in Terms of actual factors mean that all the selected various have positive or negative effect on Y1; the equations have been shown as follows ( $R^2$ = 0.9809):

Viable counts = 3.2467+1.6733\* X1-1.6683\* X3-0.5100\* X4+1.6717\* X5+0.4450\* X6+1.6567\* X7-1.5583\* X8+0.2050\* X9

Table 3. Result of ANOVA on the effect of various factors on viable counts

| Source   | SS       | DF | MS      | <b>F-Value</b> | Prob>F( p-value) |             |
|----------|----------|----|---------|----------------|------------------|-------------|
| Model    | 168.6114 | 8  | 21.0764 | 19.2157        | 0.0168           | significant |
| X1       | 33.6005  | 1  | 33.6005 | 30.6341        | 0.0116           |             |
| X3       | 33.4000  | 1  | 33.4000 | 30.4513        | 0.0117           |             |
| X4       | 3.1212   | 1  | 3.1212  | 2.8456         | 0.1902           |             |
| X5       | 33.5336  | 1  | 33.5336 | 30.5731        | 0.0117           |             |
| X6       | 2.3763   | 1  | 2.3763  | 2.1665         | 0.2374           |             |
| X7       | 32.9345  | 1  | 32.9345 | 30.0269        | 0.0120           |             |
| X8       | 29.1408  | 1  | 29.1408 | 26.5682        | 0.0142           |             |
| X9       | 0.5043   | 1  | 0.5043  | 0.4598         | 0.5463           |             |
| Residual | 3.2905   | 3  | 1.0968  |                |                  |             |
| Total    | 171.9019 | 11 |         |                |                  |             |

SS: Sum of Squares; MS: Mean Square; DF: Degree of Freedom

#### Effect of amino acids on the survival of L. bulgaricus

Table 4 shows the ANOVA of the ingredients for the survival rate of *L. bulgaricus*. The model presented a high determination coefficient ( $R^2$ = 0.9793). The Model F-value of 17.73 and p-value of 0.0188 revealed that the model is significant, there is only a 1.88% chance that a "Model F-Value" this large could occur due to noise. The relative importance of the variables was found as follows: X7> X4 > X5 > X9 > X8 > X1 > X6 > X3. Among the factors above, Arginine (X7) (p=0.0043), Glycine(X4) (p=0.0118), Leucine (X5) (p=0.0121) and Hydroxyproline (X9) (p=0.0323) can significantly affect the survival rate of *L. bulgaricus*. The linear regression equation was as follows:

# Survival rate=8.6983+0.5067\* X1+0.1817\* X3+2.7450\* X4-2.7183\* X5-0.3933 \* X6-3.9200\* X7+0.9700\* X8+1.8883\* X9

| Table 4. Result of ANO | A on the effect of various | factors on survival rate |
|------------------------|----------------------------|--------------------------|
|------------------------|----------------------------|--------------------------|

| Source   | SS       | DF | MS       | <b>F-Value</b> | Prob>F(p-value) |             |
|----------|----------|----|----------|----------------|-----------------|-------------|
| Model    | 422.9027 | 8  | 52.8628  | 17.7275        | 0.0188          | significant |
| X1       | 3.0805   | 1  | 3.0805   | 1.0331         | 0.3843          |             |
| X3       | 0.3960   | 1  | 0.3960   | 0.1328         | 0.7397          |             |
| X4       | 90.4203  | 1  | 90.4203  | 30.3224        | 0.0118          |             |
| X5       | 88.6720  | 1  | 88.6720  | 29.7361        | 0.0121          |             |
| X6       | 1.8565   | 1  | 1.8565   | 0.6226         | 0.4877          |             |
| X7       | 184.3968 | 1  | 184.3968 | 61.8373        | 0.0043          |             |
| X8       | 11.2908  | 1  | 11.2908  | 3.7864         | 0.1469          |             |
| X9       | 42.7896  | 1  | 42.7896  | 14.3495        | 0.0323          |             |
| Residual | 8.9459   | 3  | 2.9820   |                |                 |             |
| Total    | 431.8486 | 11 |          |                |                 |             |

SS: Sum of Squares; MS: Mean Square; DF: Degree of Freedom

# Effect of amino acids on the growth in medium and survival during the freezedrying of L. bulgaricus

The above-mentioned Analysis of Variance for viable counts (Y1) and survival rate (Y2) suggested that only Leucine (X5) and Arginine (X7) showed significant effect on both viability and survival. The coefficients of these two variables in linear regression equation mean that Leucine (X5) and Arginine (X7) have a positive effect on the proliferation of the cell and negative effect on the survival. Figure 1 and 2 can indicate this as the trend of the line.

### Discussion

The growth medium is a critical parameter, which is more likely to play a role in survival following freeze-drying, and the results already indicated the importance of the growth and drying medium on survival during the storage of freeze-dried *L. bulgaricus* (Carvalho *et al.*, 2004a). The effects of 16 kinds of amino acids on enriching the anti-freezing ability of *L. acidophilus* were investigated and it was found that the L-glutamic acid, L-arginine, L-leucine, L-lysine, L-methionine, L-proline, L-phenylalanine and L-threonine could promote the growth of *L. acidophilus*. L-alanine, L-isoleucine, L-cysteine, L-serine, L-phenylalanine, L-aspartic acid, glycine and L-proline could increase the anti-freezing ability of *L.* 



Figure 1. The 95% confidence interval for Leucine



Figure 2. The 95% confidence interval for Arginine

The results of the present work showed that Glutamate, Alanine, Leucine, Arginine and Lysine could affect the growth of *L. bulgaricus LB6* when added into the growth medium. However, when taking into account the survival of the cultures during freeze-drying, only a few amino acids could contribute to the survival rate of lactic acid bacteria. For instance, there were no significant differences in survival during freeze-drying after addition of sorbitol or monosodium glutamate (Carvalho *et al.*, 2003c). Kets and de Bont (1994) found that *Lactobacillus plantarum* grew significantly better in the presence of betaine under osmotically stressful conditions (0.6 M sodium chloride); however, only 11% of viable cells survived drying. Furthermore, the study has shown that glutamate, which remains inside the cell, may be responsible for the distinct survival behaviors during dehydration (Wisselink *et* 

al., 2002). Nevertheless, the present work showed that two (Leucine, Arginine) out of eight selected amino acids could significantly affect the survival of LB6, but glutamate had no effect. A study by Mattern *et al.* (1999) showed that phenylalanine, arginine, and glycine could prevent denaturation during protein vacuum drying. This could explain the protection effects of Leucine, Arginine in the present work. Therefore, the effects of these two agents on both proliferation and survival of *L. bulgaricus* when added into the growth medium were unreported in the previous studies, and the mechanism of significant effect of the two amino acids is not clear and needs further examination.

#### Conclusions

In this study, 9 selected amino acids (Glutamate, Alanine, Glycine, Leucine, Serine, Arginine, Lysine and Hydroxyproline) were investigated as promoting-growth substances in media and protective agents during freeze-drying for *L. bulgaricus LB*. Both Leucine and Arginine out of the investigated amino acids have significant effect on the growth and survival rate of *Lactobacillus bulgaricus LB*6 (p < 0.05). Moreover, they both have a positive effect on the growth and a negative effect on the survival of *L. bulgaricus LB*6.

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#### References

- Abadias, M., Benabarre, A., Teixido, N., Usall, J. 2001. Effect of freeze-drying and protectants on viability of the biocontrol yeast *Candida sake*. *International Journal of Food Microbiology*, **65**, 173–182.
- Allison, S.D., Chang, B., Randolpha, T.W., Carpentera, J.F. 1999. Hydrogen bonding between sugar and protein is responsible for inhibition of dehydration-induced protein unfolding. *Archives of Biochemistry and Biophysics*, **365**, 289-298.
- Berny, J.F. and Hennebert, G.L. 1991. Viability and stability of yeast cells and filamentous fungus spores during freeze-drying: effects of protectants and cooling rates. *Mycologia*, 83, 805–815.

- Brennan, M., Wanismail, B., Johnson, M.C., Ray, B. 1986. Cellular damage in dried Lactobacillus acidophilus. Journal of Food Protection, 49, 47-53.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P. 2003a. Effects of addition of sucrose and salt, and of starvation upon thermotolerance and survival during storage of freeze-dried *Lactobacillus delbrueckii* ssp. *bulgaricus. Journal of Food Science*, 68, 2538–2541.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P. 2003b. Effect of various growth media upon survival during storage of freeze-dried *Enterococcus faecalis* and *Enterococcus durans. Journal of Applied Microbiology*, 94,947–952.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P. 2003c. Protective effect of sorbitol and monosodium glutamate during storage of freeze-dried lactic acid bacteria. *Dairy Science and Technology*. 83, 203–210.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P. 2004a. Effects of various sugars added to growth and drying media upon thermotolerance and survival throughout storage of freeze-dried *Lactobacillus delbrueckii* ssp. *bulgaricus.*" *Biotechnology Progress*, 20, 248–254.
- Carvalho, A.S., Silva, J., Ho, P., Teixeira, P., Malcata, F.X., Gibbs, P. 2004b. Relevant factors for the preparation of freeze-dried lactic acid bacteria. *International Dairy Journal*, 14, 835–847.
- Chen, H., Chen, S. W., Chen, H. L., Wu, Y. Y., Shu, G. W. 2014. Effects of carbon sources and prebiotics added to growth media on proliferation and survival of *Lactobacillus bulgaricus* LB6 during freeze-drying. *Journal of Chemical and Pharmaceutical Research*, 6(6), 894-899.
- Chen, H, Chen S.W., Chen H.L. Wu Y.Y., Shu, G.W. 2015. Effects of sugar alcohol and proteins on the survival of *Lactobacillus bulgaricus* LB6 during freeze-drying. *Acta Scientiarum Polonorum Technologia Alimentaria*, 14(2), 117–124
- Depaz, R.A., Dale, D.A., Barnett, C.C., Carpenter, J.F. 2002. Effects of drying methods and additives on the structure, function, and storage stability of subtilisin; role of protein conformation and molecular mobility. *Enzyme and Microbial Technology*, **31**, 765-774.
- Efiuvwevwere, B.J., Gorris, L.G., Smid, E.J., Kets, E.P. 1999. Mannitol-enhanced survival of *Lactococcus lactis* subjected to drying. *Applied Microbiology and Biotechnology*, **51**, 100–104.
- Ferreira, V., Soares, V., Santos, C., Silva, J., Gibbs, P. A. 2005. Survival of *Lactobacillus sakei* during heating, drying and storage in the dried state when growth has occurred in the presence of sucrose or monosodium glutamate. *Biotechnology Letters*, 27, 249–252.

- Foerst, P., Kulozik, U., Schmitt, M., Bauer, S., Santivarangkna, S. 2012. Stability of vacuumdried probiotic bacterium *Lactobacillus paracasei* F19. *Food and Bioproducts Processing*, **90**, 295–300.
- Fonseca,F., Bèal, C., Corrieu, G. 2000. Method for quantifying the loss of acidification activity of lactic acid starters during freezing and frozen storage. *Journal of Dairy Research*, 67, 83–90.
- Font, V. G., Savoy G.G., Pesce R.H., Oliver, G.A. 1983. Comparative study of the efficiency of some additives in protecting lactic acid bacteria against freeze-drying. *Cryobiology*. 20, 560-566.
- Fowler, A. and Toner, M. 2005.Cryo-injury and biopreservation. Annals of the New York Academy of Sciences, 1066, 119–135.
- Guchte, M., Penaud, S., Grimaldi, C., Barbe, V. 2006. The complete genome sequence of Lactobacillus bulgaricus reveals extensive and ongoing reductive evolution. Proceedings of the National Academy of Sciences, 103, 9274–9279.
- Higl, B., Kurtmann, L., Carlsen, C.U., Ratjen, J. 2007. Impact of water activity, temperature, and physical state on the storage stability of *Lactobacillus paracasei* ssp. *paracasei* freeze-dried in a lactose matrix. *Biotechnology Progress*, 23, 794–800.
- Hubalek, Z. 2003. Protectants used in the cryopreservation of microorganisms. *Cryobiology*. 46, 205–229.
- Kets, E.P. and Bont, J.A. 1994. Protective effect of betaine on survival of subjected to drying. *FEMS Microbiology Letters*, **116**, 251–256.
- Kets, E.P., Jpelaar, P.J., Hoekstra, F.A., Vromansa, H. 2004. Citrate increases glass transition temperature of vitrified sucrose preparations. *Cryobiology*. 48, 46-54.
- Kurtmann, L., Carlsen, C.U., Risbo, J., Skibsted, L.H. 2009. Storage stability of freeze–dried *Lactobacillus acidophilus* (La-5) in relation to water activity and presence of oxygen and ascorbate. *Cryobiology*. 58, 175–180.
- Linders, L.J.M., de Jong, G.I.W., Meerdink, G., van't Riet, K. 1997a. Carbohydrates and the dehydration inactivation of *Lactobacillus plantarum*: the role of moisture distribution and water activity. *Journal of Food Engineering*, **31**, 237–250.
- Linders, L.J.M., Wolkers, W.F., Hoekstra, F.A., van't Rieta, K. 1997b. Effect of added carbohydrates on membrane phase behavior and survival of dried *Lactobacillus plantarum*. *Cryobiology*. **35**, 31–40.
- Lone K.C., Carlsen, J.R., Skibsted, L.H. Storage stability of freeze-dried *Lactobacillus* acidophilus (La-5) in relation to water activity and presence of oxygen and ascorbate.

Cryobiology 2009, 58(2), 175-180.

- Mattern, M., Winter, G., Kohnert, U., Lee, G. 1999. Formulation of proteins in vacuum-dried glasses. II. Process and storage stability in sugar-free amino acid systems. *Pharmaceutical Development and Technology*, 4(2), 199-208.
- Meng, X.C., Stanton, C., Fitzgerald, G.F., Daly, C., Ross, R.P. 2008. Anhydrobiotics: The challenges of drying probiotic cultures. *Food Chemistry*, **106**, 1406-1416.
- Morgan, C., and Vesey, G. 2009. Freeze Drying of Microorganisms Encyclopedia of Microbiology. Third Edition, 162–173.
- Ohtake, S. 2004. Effect of sugar-phosphate mixtures on the stability of DPPC membranes in dehydrated systems. *Cryobiology*, 48, 81-89.
- Passot, S., Cenard, S., Douania, I., Tréléa, I.C. 2012. Critical water activity and amorphous state for optimal preservation of lyophilised lactic acid bacteria. *Food Chemistry*, **132** (4), 1699–1705.
- Santivarangkna, C., Higl, B., Foerst, P. 2008. Protection mechanisms of sugars during different stages of preparation process of dried lactic acid starter cultures. *Food Microbiology*, 25, 429–441.
- Shu, G.W., Li, C.N., Chen, H. Wang, C.F. 2014. Effect of inoculum and temperature on the fermentation of goat yogurt. *Advance Journal of Food Science and Technology*, 6, 68-71.
- Teixeira, P.C., Castro, M.H., Malcata, F.X., Kirby, R.M. 1995. Survival of Lactobacillus delbruckii ssp. bulgaricus following spray drying. Journal of Dairy Science, 78, 1025– 1031.
- Wang L., Chen H., Shu, G.W., Shi J.F., Chen G. Effects of amino acids on the growth and freeze-drying of *Lactobacillus acidophilus*. China brewing, 2011, **30**(2), 59-62.
- Wisselink, H. W., Weusthuis, R. A., Eggink, G., Hugenholtz, J., Grobben, G. J., 2002. Mannitol production by lactic acid bacteria: A review. *International Dairy Journal*, **12**, 151–161.